

**Small scale collaborative trial on the method for the
determination of Metolachlor in TC, EC and EW**

5318/R

Report to CIPAC
By
Chinese Pesticide Analytical Committee (CHIPAC)

Method Developed by Shandong Binnong Technology Co., Ltd

05/2022

Table of Content

1. Metolachlor method description	3
2. Participants and sample distribution	5
3. Deviations and remarks.....	6
4. Statistical evaluation.....	6
5. Conclusion.....	17
6. Figures.....	19

Small scale collaborative trial on the methods for the determination of metolachlor in TC, EC and EW

1. Metolachlor method description

1. 1 Outline of method

Metolachlor is dissolved in methanol and determined by gas chromatography with flame ionization detection, using dipentyl phthalate as internal standard.

1. 2 Apparatus and reagents

GC system with FID

Column, HP-5, capillary, 30 m x 0.32 mm×0.25µm, or equivalent.

Electronic integrator or data system

ultrasonic bath

Nylon syringe filters, 0.22 µm

Metolachlor standard of known purity

Dipentyl phthalate pure, internal standard. Should not contain any impurities that elute at the metolachlor retention time.

Methanol, HPLC grade

Internal standard solution. Weigh into a volumetric flask (250 ml) dipentyl phthalate (2.5 g). Add methanol (200 ml) into the flask, place the flask in an ultrasonic bath for 2-3 min. Allow to cool to ambient temperature. Dilute to volume with methanol. Mix thoroughly.

Calibration solutions. Weigh in duplicate (to the nearest 0.1 mg) into a volumetric flask (10 ml) about 50 mg of metolachlor standard (s mg). Add internal standard solution (5 ml) into the flask, place the flask in an ultrasonic bath for 2-3 min. Allow to cool to ambient temperature. Dilute to volume with methanol. Mix thoroughly. Filter a portion of each sample solution with a 0.22µm filter prior to analysis (Solutions CA and CB).

1. 3 GC condition

Column temperature 182°C

Injection port temperature	250°C
Detector temperature	250°C
Injection volume	1.0 µl
Flow rate	
nitrogen (carrier)	2.0 ml/min
hydrogen	30 ml/min
air	300 ml/min
make up	25 ml/min
Injection type	Split
Split ratio	20:1
Retention time	Metolachlor: about 8.6 min internal standard: about 15.2 min

1.4 Procedure

(i) Preparation of Metolachlor sample. Weigh in duplicate (to the nearest 0.1 mg) into a volumetric flask (10 ml) sufficient sample to contain about 50 mg of metolachlor (w mg). Add internal standard solution (5 ml) into the flask, place the flask in an ultrasonic bath for 2-3 min. Allow to cool to ambient temperature. Dilute to volume with methanol. Mix thoroughly. Filter a portion of each sample solution with a 0.22µm filter prior to analysis (Solutions S1 and S2).

(ii) Determination of metolachlor

Inject into the gas chromatograph 1µl portions of the calibration solution until the area (or height) ratios (R') of metolachlor to the internal standard varies by less than 1.5% for successive injections, otherwise prepare new calibration solutions. Inject in duplicate 1µl portions of each sample solution bracketing them by injections of the calibration solutions as follows: CA, S1, S1, CB, S2, S2, CA, and so on.

(iii) Calculation

Calculate the mean value of each pair of response factors bracketing the two injections of a sample and use this value for calculating the metolachlor contents of the bracketed

sample injections. The metolachlor content is the mean value of two sample solutions.

$$f_i = \frac{s \times P}{R'}$$

$$\text{Metolachlor content} = \frac{R \times f}{w} \text{ g/kg}$$

where:

R = peak area (or height) ratio of metolachlor to the internal standard for the sample solution

R' = peak area (or height) ratio of metolachlor to the internal standard for the calibration solution

f_i = individual response factor

f = mean response factor

s = mass of metolachlor standard taken (mg)

w = mass of sample taken (mg)

P = purity of the standard metolachlor (g/kg)

2. Participants and sample distribution

Participants

Index	NAME	ORGANIZATION	Address
Lab1	Dong Jie	Shandong Binnong Technology Co., Ltd.	No.518, Yongxin Road, Binbei Town, Binzhou, Shandong, China
Lab2	Shao Xiangdong	BioGuide Technologies Co., Ltd	Building 8, IFST-CAAS, 2 Yuanmingyuan West Road, Haidian District, Beijing 100193, China.
Lab3	Sun Hongfeng	China National Pesticide Quality Supervision Testing Center (Shenyang)	No.8, Shenliao East Road, Tiexi District, Shenyang City, P.R. of China

Sample information

Sample	Quantity	Batch	Declared Content of AI
Metolachlor TC-1	100 g	202106034	980 g/kg
Metolachlor TC-2	100 g	202107046	980 g/kg
Metolachlor TC-3	100 g	202108058	980 g/kg
Metolachlor EC-1	100 mL	202107048	670 g/kg
Metolachlor EC-2	100 mL	202107134	670 g/kg
Metolachlor EC-3	100 mL	202108069	670 g/kg
Metolachlor EW-1	100 mL	202106025	500 g/kg
Metolachlor EW-2	100 mL	202107042	500 g/kg
Metolachlor EW-3	100 mL	202108043	500 g/kg

3. Deviations and remarks

Lab 2 used a flow rate of 1.9 mL/m and column temperature of 181 °C.

Column information:	HP-5, 30m×0.32mm×0.25μm	DB-5, 30m×0.32mm×0.25μm	HP-5 30m×0.32mm×0.25μm
Column Temperature (°C):	182	181	182
Injection port temperature (°C):	250	250	250
Detector temperature (°C):	250	250	250
Flow rate - nitrogen (carrier), ml/min	2.0	1.9	2.0
Flow rate - hydrogen, ml/min	30	30	30
Flow rate - air, ml/min	300	300	300
Flow rate - make up, ml/min	25	25	25
Injection Volume (μL):	1.0	1	1
Split ratio	20:1	20:1	20:1

4. Statistical evaluation

Table 1. Results of the analysis of Al content in the TC-1

Lab	Day1 (g/kg)		Day2 (g/kg)		Average Yi(g/kg)	Yi ²	Standard Deviation Si	Si ²
	1	2	1	2				
Lab1	980.5	986.5	981.1	984.8	983.2	966756.0	2.9136	8.4890
Lab2	975.9	974.4	973.9	978.6	975.7	951980.7	2.1140	4.4689
Lab3	976.7	969.7	975.0	975.8	974.3	949226.4	3.1162	9.7105

Table 2. Results of the analysis of Al content in the TC-2

Lab	Day1 (g/kg)		Day2 (g/kg)		Average Yi(g/kg)	Yi ²	Standard Deviation n Si	Si ²
	1	2	1	2				
Lab1	978.93	981.06	982.25	980.86	980.8	961919.6006	1.3745	1.8894
Lab2	974.47	973.85	974.11	973.57	974.0	948676.0000	0.3831	0.1468
Lab3	973.61	970.47	974.37	972.84	972.8	946383.6165	1.6881	2.8498

Table 3. Results of the analysis of Al content in the TC-3

Lab	Day1 (g/kg)		Day2 (g/kg)		Average Yi(g/kg)	Yi ²	Standard Deviation Si	Si ²
	1	2	1	2				

Lab1	980.19	981.32	981.44	984.42	981.8	964014.6948	1.8082	3.2698
Lab2	981.46	984.52	977.96	971.62	978.9	958225.6321	5.5383	30.6732
Lab3	971.68	971.21	971.23	971.86	971.5	943802.5350	0.3260	0.1063

Table 4. Results of the analysis of Al content in the EC-1

Lab	Day1 (g/kg)		Day2 (g/kg)		Average Yi(g/kg)	Yi ²	Standard Deviation Si	Si ²
	1	2	1	2				
Lab1	676.0	676.6	673.6	676.2	675.6	456428.6	1.3663	1.8668
Lab2	682.9	692.0	688.8	681.8	686.4	471086.6	4.8640	23.6582
Lab3	670.7	674.4	670.7	680.3	674.0	454269.3	4.5270	20.4936

Table 5. Results of the analysis of Al content in the EC-2

Lab	Day1 (g/kg)		Day2 (g/kg)		Average Yi(g/kg)	Yi ²	Standard Deviation Si	Si ²
	1	2	1	2				
Lab1	674.4	673.3	669.7	676.9	673.6	453676.3380	2.9920	8.9523
Lab2	686.7	681.7	687.1	683.6	684.8	468906.5291	2.5978	6.7484
Lab3	674.9	674.7	677.1	674.9	675.4	456131.3906	1.1339	1.2858

Table 6. Results of the analysis of Al content in the EC-3

Lab	Day1 (g/kg)		Day2 (g/kg)		Average Yi(g/kg)	Yi ²	Standard Deviation Si	Si ²
	1	2	1	2				
Lab1	675.9	672.5	671.0	674.6	673.5	453595.5150	2.1777	4.7424
Lab2	691.5	684.0	688.7	681.1	686.3	471059.1639	4.6623	21.7366
Lab3	674.9	679.9	678.3	680.0	678.3	460046.8016	2.4075	5.7959

Table 7. Results of the analysis of Al content in the EW-1

Lab	Day1 (g/kg)		Day2 (g/kg)		Average Yi(g/kg)	Yi ²	Standard Deviation Si	Si ²
	1	2	1	2				
Lab1	493.2	494.3	501.4	498.3	496.8	246817.6921	3.7524	14.0805
Lab2	504.3	501.8	493.8	494.6	498.6	248616.9182	5.2146	27.1923
Lab3	492.4	491.4	498.5	495.8	494.5	244542.6127	3.2874	10.8067

Table 8. Results of the analysis of Al content in the EW-2

Lab	Day1 (g/kg)		Day2 (g/kg)		Average Yi(g/kg)	Yi ²	Standard Deviation	Si ²

	1	2	1	2			Si	
Lab1	486.4	483.1	487.5	486.0	485.7	235945.7763	1.8494	3.4202
Lab2	490.3	490.7	482.8	485.7	487.4	237531.9538	3.7783	14.2755
Lab3	483.9	476.8	484.8	484.0	482.4	232690.4644	3.7367	13.9627

Table 9. Results of the analysis of Al content in the EW-3

Lab	Day1 (g/kg)		Day2 (g/kg)		Average Yi(g/kg)	Yi ²	Standard Deviation Si	Si ²
	1	2	1	2				
Lab1	492.6	494.3	494.8	494.0	493.9	243964.3753	0.9336	0.8717
Lab2	499.4	493.2	490.1	495.7	494.6	244616.7952	3.9402	15.5251
Lab3	485.9	486.2	492.3	492.0	489.1	239211.4736	3.5067	12.2966

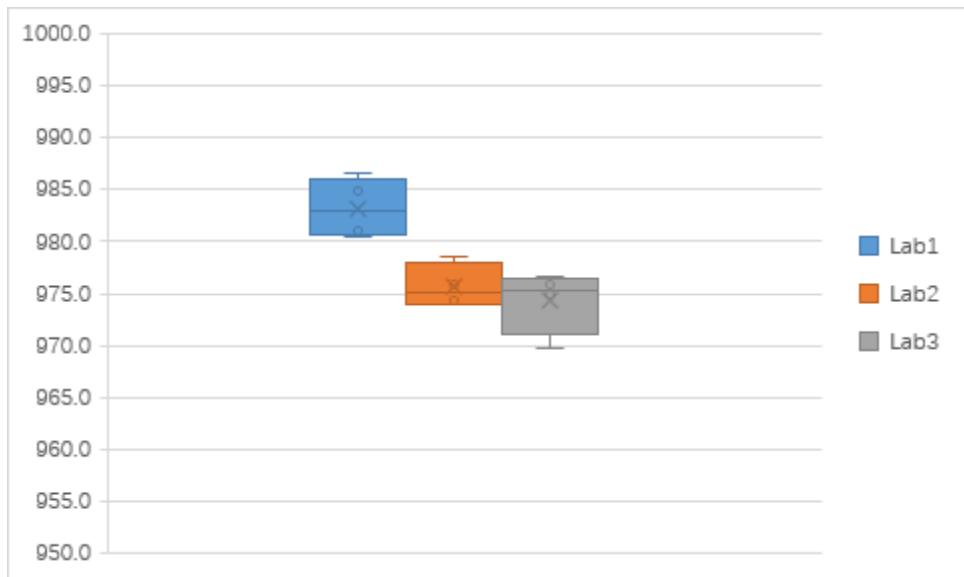


Figure 1. Graphical presentation of TC-1 data

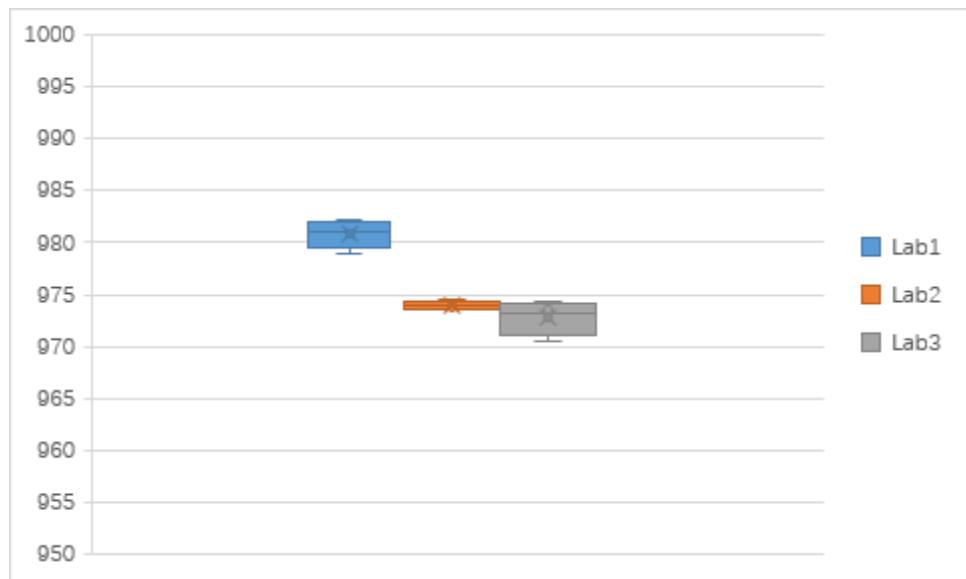


Figure 2. Graphical presentation of TC-2 data



Figure 3. Graphical presentation of TC-3 data

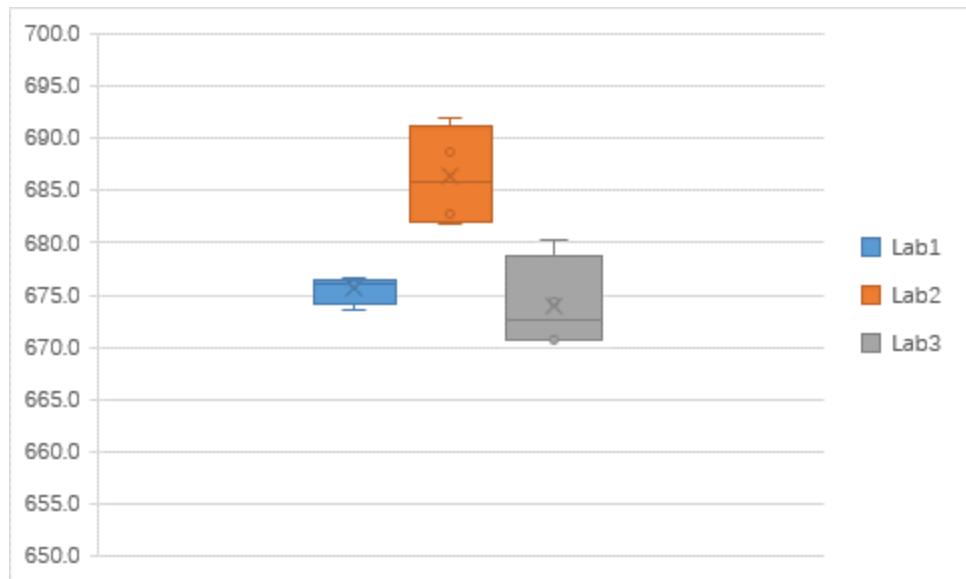


Figure 4. Graphical presentation of EC-1 data

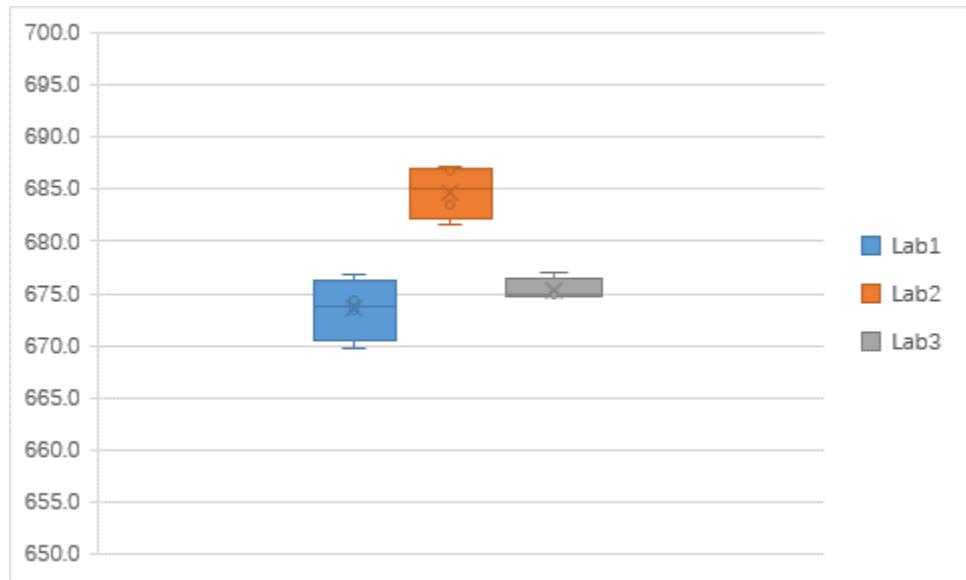


Figure 5. Graphical presentation of EC-2 data

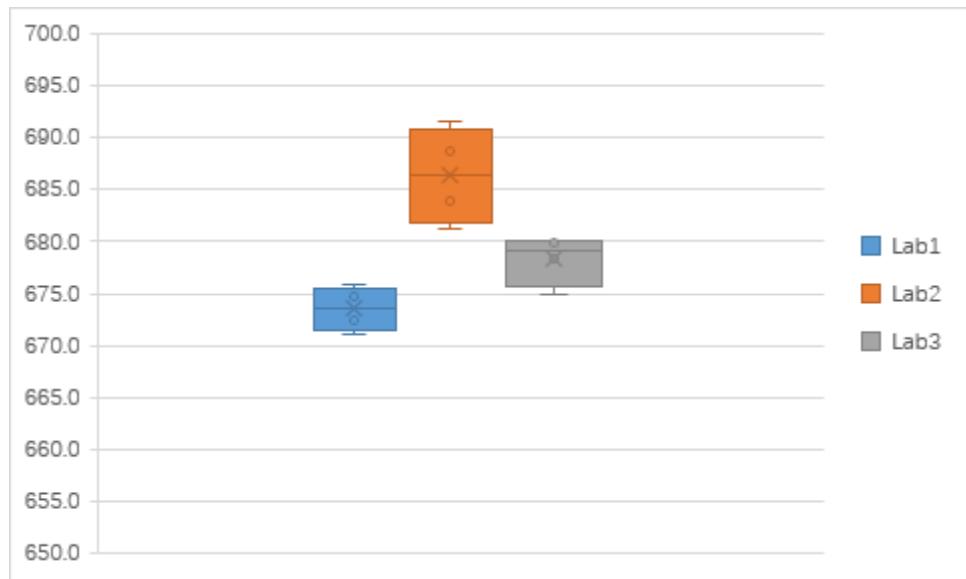


Figure 6. Graphical presentation of EC-3 data



Figure 7. Graphical presentation of EW-1 data

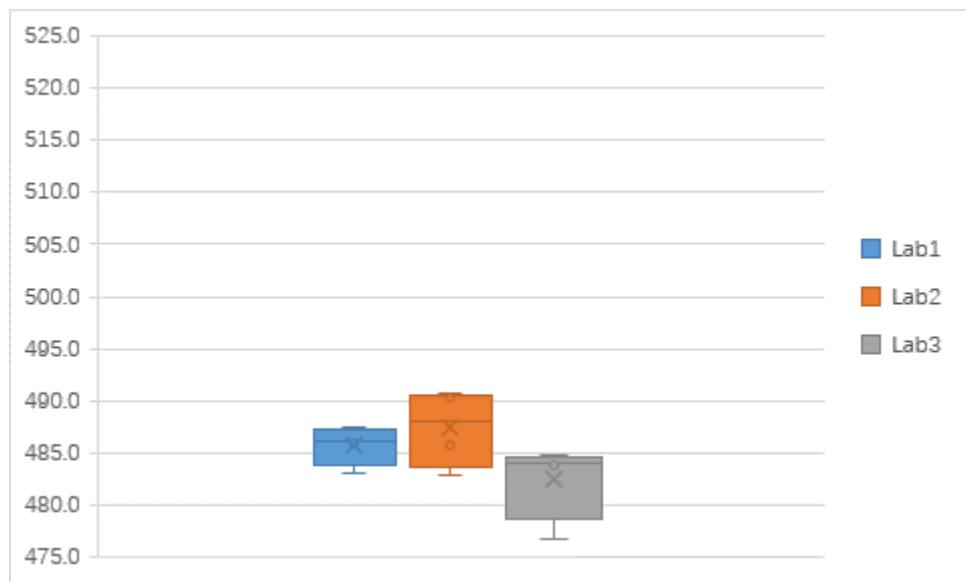


Figure 8. Graphical presentation of EW-2 data

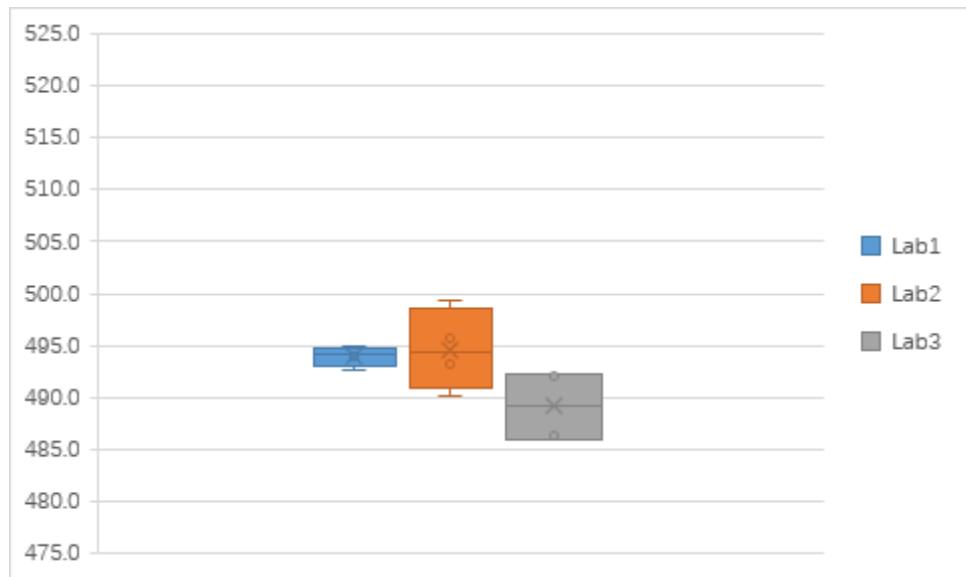


Figure 9. Graphical presentation of EW-3 data

Table 10. Statistics of the results of TC-1

$S_1 = \text{sum } Y_i$	2933.22		
$S_2 = \text{sum } Y_i^2$	2867963.10424		
$S_3 = \text{sum } S_i^2$	22.6683		
No. Lab P	3		
No. Determination n	4		
Average $Y = S_1/P$	977.74		
$S_r^2 = S_3/P$	7.5561	Standard Deviation of Repeatability S_r	2.7488
$S_L^2 = [(P \cdot S_2 - S_1^2)/P(P-1)] - S_r^2/n$	21.2904	S_L	4.6142

$S_r^2 = S_r^2 + S_L^2$	28.8465	Standard Deviation Reproducibility S_R	5.3709
Repeatability $r=2.8 \cdot S_r$	7.6967		
Reproducibility $R=2.8 \cdot S_R$	15.0385		
Relative Standard Deviation of Repeatability $RSD_r = S_r \cdot 100/Y$	0.2811		
Relative Standard Deviation of Reproducibility $RSD_R = S_R \cdot 100/Y$	0.5493		
Horwitz RSD_R $(Hor)=2^{[1-0.5 \cdot \log(Y/1000)]}$	2.0068		
HorRat	0.27372994		

Table 11. Statistics of the results of TC-2

$S_1 = \text{sum } Y_i$	2927.60		
$S_2 = \text{sum } Y_i^2$	2856979.21713		
$S_3 = \text{sum } S_i^2$	4.8860		
No. Lab P	3		
No. Determination n	4		
Average $Y = S_1/P$	975.87		
$S_r^2 = S_3/P$	1.6287	Standard Deviation of Repeatability S_r	1.2762
$S_L^2 = [(P \cdot S_2 - S_1^2)/P(P-1)] - S_r^2/n$	18.0144	S_L	4.2443
$S_R^2 = S_r^2 + S_L^2$	19.6431	Standard Deviation Reproducibility S_R	4.4320
Repeatability $r=2.8 \cdot S_r$	3.5733		
Reproducibility $R=2.8 \cdot S_R$	12.4097		
Relative Standard Deviation of Repeatability $RSD_r = S_r \cdot 100/Y$	0.1308		
Relative Standard Deviation of Reproducibility $RSD_R = S_R \cdot 100/Y$	0.4542		
Horwitz RSD_R $(Hor)=2^{[1-0.5 \cdot \log(Y/1000)]}$	2.0074		
HorRat	0.226249456		

Table 12. Statistics of the results of TC-3

$S_1 = \text{sum } Y_i$	2932.23		
$S_2 = \text{sum } Y_i^2$	2866042.86193		
$S_3 = \text{sum } S_i^2$	34.0493		
No. Lab P	3		
No. Determination n	4		

Average Y=S ₁ /P	977.41		
S _r ² =S ₃ /P	11.3498	Standard Deviation of Repeatability S _r	3.3689
S _L ² =[(P*S ₂ -S ₁ ²)/P(P-1)]- S _r ² /n	25.5749	S _L	5.0572
S _R ² =S _r ² +S _L ²	36.9247	Standard Deviation Reproducibility S _R	6.0766
Repeatability r=2.8*S _r	9.4330		
Reproducibility R=2.8*S _R	17.0144		
Relative Standard Deviation of Repeatability RSD _r =S _r *100/Y	0.3447		
Relative Standard Deviation of Reproducibility RSD _R =S _R *100/Y	0.6217		
Horwitz RSD _R (Hor)=2^-[1-0.5*log(Y/1000)]	2.0069		
HorRat	0.309783417		

Table 13. Statistics of the results of EC-1

S ₁ =sum Yi	2035.95		
S ₂ =sum Yi ²	1381784.48186		
S ₃ =sum Si ²	46.0186		
No. Lab P	3		
No. Determination n	4		
Average Y=S ₁ /P	678.65		
S _r ² =S ₃ /P	15.3395	Standard Deviation of Repeatability S _r	3.9166
S _L ² =[(P*S ₂ -S ₁ ²)/P(P-1)]- S _r ² /n	41.3689	S _L	6.4319
S _R ² =S _r ² +S _L ²	56.7085	Standard Deviation Reproducibility S _R	7.5305
Repeatability r=2.8*S _r	10.9664		
Reproducibility R=2.8*S _R	21.0854		
Relative Standard Deviation of Repeatability RSD _r =S _r *100/Y	0.5771		
Relative Standard Deviation of Reproducibility RSD _R =S _R *100/Y	1.1096		
Horwitz RSD _R (Hor)=2^-[1-0.5*log(Y/1000)]	2.1202		
HorRat	0.52336985		

Table 14. Statistics of the results of EC-2

$S_1=\text{sum } Y_i$	2033.70		
$S_2=\text{sum } Y_i^2$	1378714.25771		
$S_3=\text{sum } S_i^2$	16.9865		
No. Lab P	3		
No. Determination n	4		
Average $Y=S_1/P$	677.90		
$S_r^2=S_3/P$	5.6622	Standard Deviation of Repeatability S_r	2.3795
$S_L^2=[(P*S_2-S_1^2)/P(P-1)] - S_r^2/n$	34.7931	S_L	5.8986
$S_R^2=S_r^2+S_L^2$	40.4552	Standard Deviation Reproducibility S_R	6.3604
Repeatability $r=2.8*S_r$	6.6627		
Reproducibility $R=2.8*S_R$	17.8092		
Relative Standard Deviation of Repeatability $RSD_r=S_r*100/Y$	0.3510		
Relative Standard Deviation of Reproducibility $RSD_R=S_R*100/Y$	0.9383		
Horwitz RSD_R $(Hor)=2^{[1-0.5*\log(Y/1000)]}$	2.1205		
HorRat	0.442466128		

Table 15. Statistics of the results of EC-3

$S_1=\text{sum } Y_i$	2038.10		
$S_2=\text{sum } Y_i^2$	1384701.48049		
$S_3=\text{sum } S_i^2$	32.2749		
No. Lab P	3		
No. Determination n	4		
Average $Y=S_1/P$	679.37		
$S_r^2=S_3/P$	10.7583	Standard Deviation of Repeatability S_r	3.2800
$S_L^2=[(P*S_2-S_1^2)/P(P-1)] - S_r^2/n$	39.4490	S_L	6.2808
$S_R^2=S_r^2+S_L^2$	50.2073	Standard Deviation Reproducibility S_R	7.0857
Repeatability $r=2.8*S_r$	9.1840		
Reproducibility $R=2.8*S_R$	19.8400		
Relative Standard Deviation of Repeatability $RSD_r=S_r*100/Y$	0.4828		

Relative Standard Deviation of Reproducibility $RSD_R = S_R * 100/Y$	1.0430		
Horwitz RSD_R $(Hor) = 2^{[1 - 0.5 \cdot \log(Y/1000)]}$	2.1198		
HorRat	0.492015105		

Table 16. Statistics of the results of EW-1

$S_1 = \text{sum } Y_i$	1489.94		
$S_2 = \text{sum } Y_i^2$	739977.22294		
$S_3 = \text{sum } S_i^2$	52.0795		
No. Lab P	3		
No. Determination n	4		
Average $Y = S_1/P$	496.65		
$S_r^2 = S_3/P$	17.3598	Standard Deviation of Repeatability S_r	4.1665
$S_L^2 = [(P \cdot S_2 - S_1^2)/P(P-1)] - S_r^2/n$	0.1125	S_L	0.3354
$S_R^2 = S_r^2 + S_L^2$	17.4724	Standard Deviation Reproducibility S_R	4.1800
Repeatability $r = 2.8 \cdot S_r$	11.6662		
Reproducibility $R = 2.8 \cdot S_R$	11.7040		
Relative Standard Deviation of Repeatability $RSD_r = S_r * 100/Y$	0.8389		
Relative Standard Deviation of Reproducibility $RSD_R = S_R * 100/Y$	0.8416		
Horwitz RSD_R $(Hor) = 2^{[1 - 0.5 \cdot \log(Y/1000)]}$	2.2222		
HorRat	0.378747709		

Table 17. Statistics of the results of EW-2

$S_1 = \text{sum } Y_i$	1455.50		
$S_2 = \text{sum } Y_i^2$	706168.19446		
$S_3 = \text{sum } S_i^2$	31.6584		
No. Lab P	3		
No. Determination n	4		
Average $Y = S_1/P$	485.17		
$S_r^2 = S_3/P$	10.5528	Standard Deviation of Repeatability S_r	3.2485
$S_L^2 = [(P \cdot S_2 - S_1^2)/P(P-1)] - S_r^2/n$	3.8432	S_L	1.9604

$S_r^2 = S_r^2 + S_L^2$	14.3960	Standard Deviation Reproducibility S_R	3.7942
Repeatability $r=2.8 \cdot S_r$	9.0958		
Reproducibility $R=2.8 \cdot S_R$	10.6238		
Relative Standard Deviation of Repeatability $RSD_r = S_r \cdot 100/Y$	0.6696		
Relative Standard Deviation of Reproducibility $RSD_R = S_R \cdot 100/Y$	0.7820		
Horwitz RSD_R ($Hor = 2^{[1-0.5 \cdot \log(Y/1000)]}$)	2.2300		
HorRat	0.350689859		

Table 18. Statistics of the results of EW-3

$S_1 = \sum Y_i$	1477.61		
$S_2 = \sum Y_i^2$	727792.64397		
$S_3 = \sum S_i^2$	28.6934		
No. Lab P	3		
No. Determination n	4		
Average $Y = S_1/P$	492.54		
$S_r^2 = S_3/P$	9.5645	Standard Deviation of Repeatability S_r	3.0926
$S_L^2 = [(P \cdot S_2 - S_1^2)/P(P-1)] - S_r^2/n$	6.6102	S_L	2.5710
$S_R^2 = S_r^2 + S_L^2$	16.1747	Standard Deviation Reproducibility S_R	4.0218
Repeatability $r=2.8 \cdot S_r$	8.6594		
Reproducibility $R=2.8 \cdot S_R$	11.2610		
Relative Standard Deviation of Repeatability $RSD_r = S_r \cdot 100/Y$	0.6279		
Relative Standard Deviation of Reproducibility $RSD_R = S_R \cdot 100/Y$	0.8165		
Horwitz RSD_R ($Hor = 2^{[1-0.5 \cdot \log(Y/1000)]}$)	2.2250		
HorRat	0.366992373		

A Grubbs test were run on all determinations, and no outlier was found.

5. Conclusion

No outlier was found in the data set.

All RSD_R values were lower than the criteria calculated using the Horwitz equation. All HorRat values were smaller than 1.0, while 2 values fell below 0.3.

From the results shown above, the method can be considered applicable for the determination of metolachlor contents in TC, EC and EW. CHIPAC propose that a full scale collaborative trial might be conducted.

6. Figures

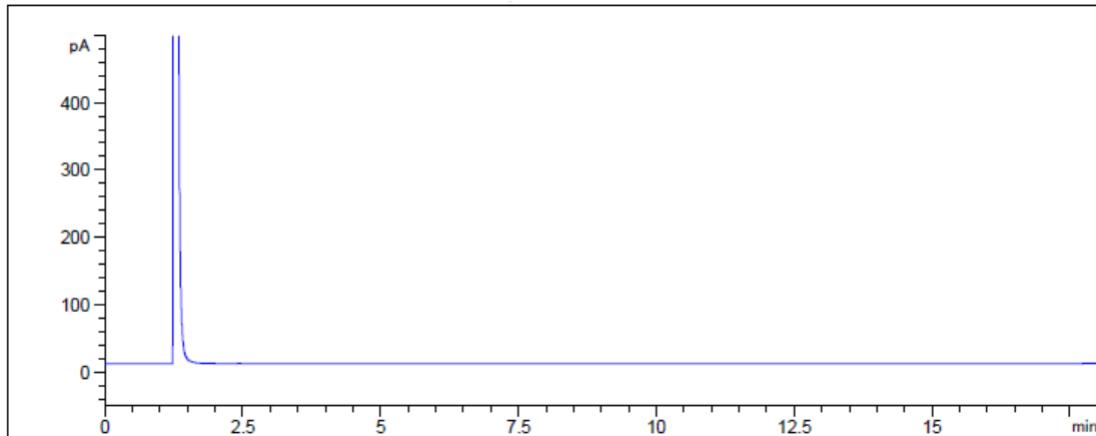


Fig.1 GC chromatogram of solvent blank

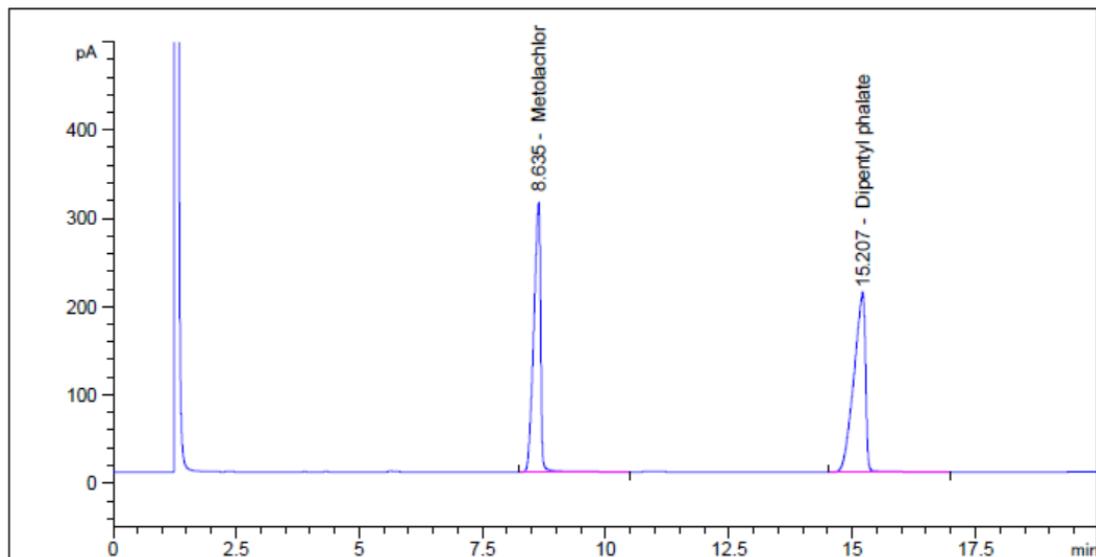


Fig.2 GC chromatogram of Metolachlor standard

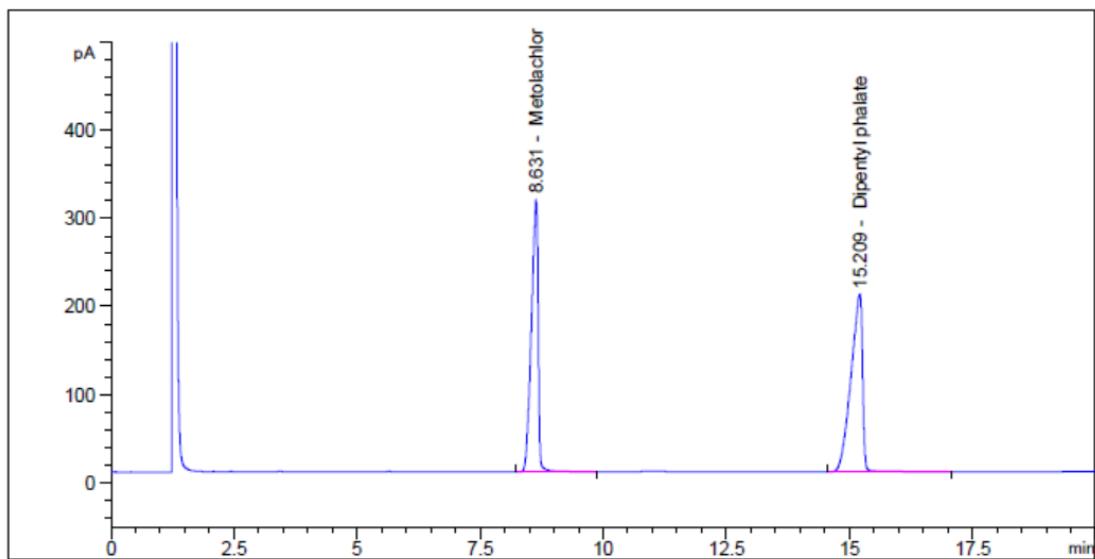


Fig.3 GC chromatogram of Metolachlor TC

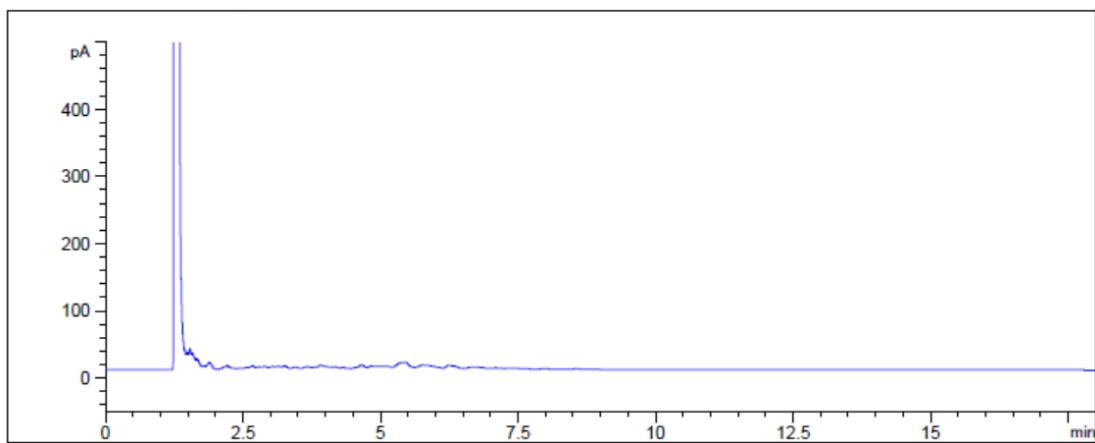


Fig.4 GC chromatogram of Metolachlor EC blank

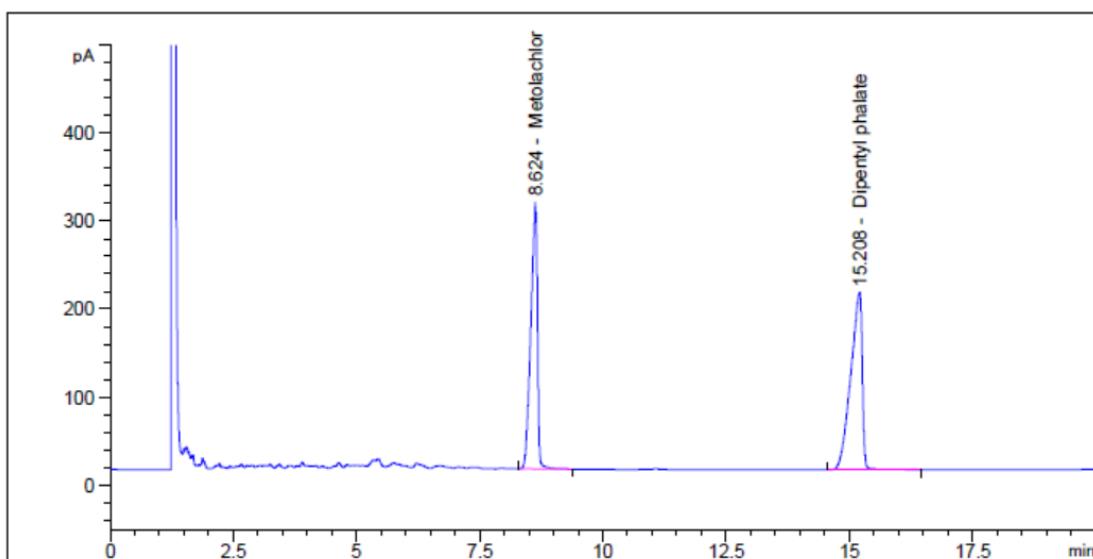


Fig.5 GC chromatogram of Metolachlor EC

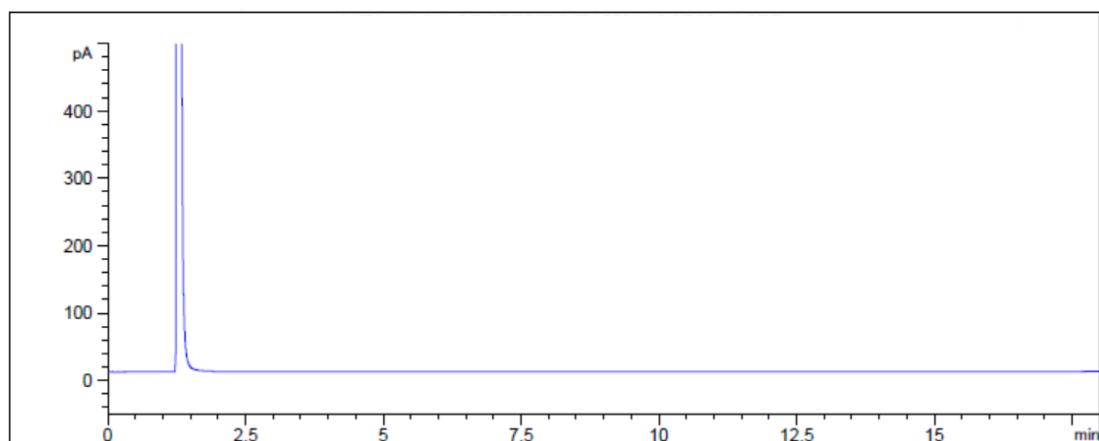


Fig.6 GC chromatogram of Metolachlor EW blank

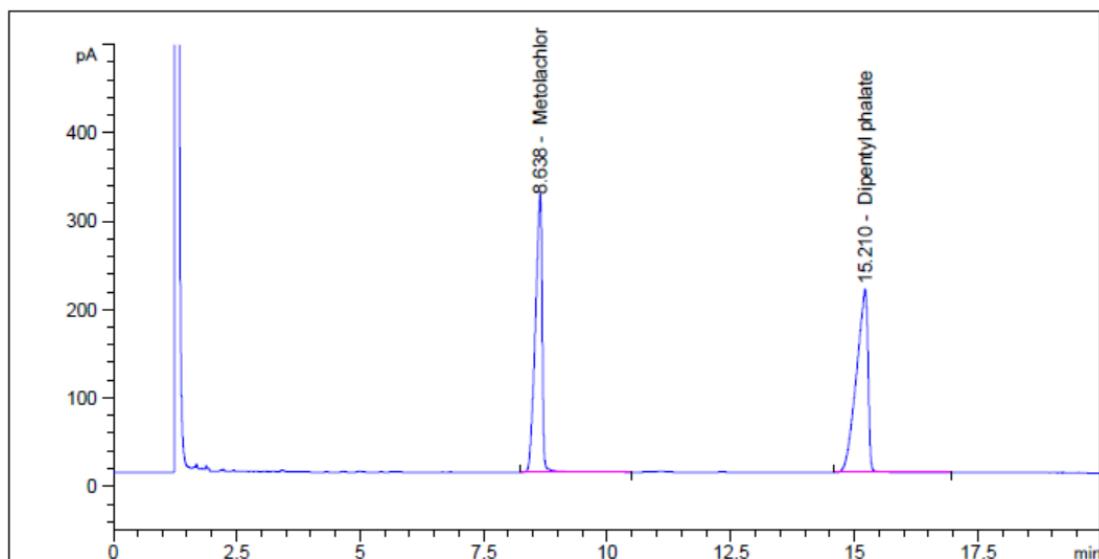


Fig.7 GC chromatogram of Metolachlor EW